

## Note

# Efficient syntheses of 1-bromodeoxy-, 1-azidodeoxy- and 1-aminodeoxypentitols from unprotected D-pentono-1,4-lactones

Véronique Bouchez, Imane Stasik \*, Daniel Beaupère

*Laboratoire de Chimie Organique, Université de Picardie Jules Verne, 33 rue Saint-Leu, F-80039 Amiens, France*

Received 7 July 1999; accepted 15 September 1999

## Abstract

The reduction of unprotected 5-bromo-5-deoxy-D-ribo-, D-arabinono and D-xylono-1,4-lactones was achieved with NaBH<sub>4</sub> in water–EtOH. The corresponding 1-bromo-1-deoxypentitols were isolated after acetylation in good overall yields (60–90%). 1-Azido-1-deoxypentitols were obtained quantitatively either by nucleophilic substitution by azide ion and deacetylation of the corresponding monobromopentitols or by reduction of the corresponding 5-azido-5-deoxy-D-pentono-1,4-lactones. The reduction of the monoazidopentitols by catalytic hydrogen transfer gave the monoaminopentitol analogues in quantitative yield. © 2000 Elsevier Science Ltd. All rights reserved.

**Keywords:** Unprotected D-pentono-1,4-lactones; Reduction; Monobromoalditols; Monoazidoalditols; Monoaminoalditols

## 1. Introduction

Monoaminodeoxyalditols are used as intermediates for the synthesis of their anhydrides [1], which are useful synthons for the synthesis of nucleoside analogues [2,3], and as potential enzyme inhibitors [4–6]. Reductive amination of aldoses with benzylamine–sodium borohydride and subsequent catalytic hydrogenation gave the corresponding 1-amino-1-deoxyalditol hydrochlorides in 39–68% overall yield [1]. To our knowledge, most previously reported 1-amino-1-deoxyalditols belong to the D-series, with 1-amino-1-deoxy-L-rhamnitols [1] being the only example from the L-series.

## 2. Results and discussion

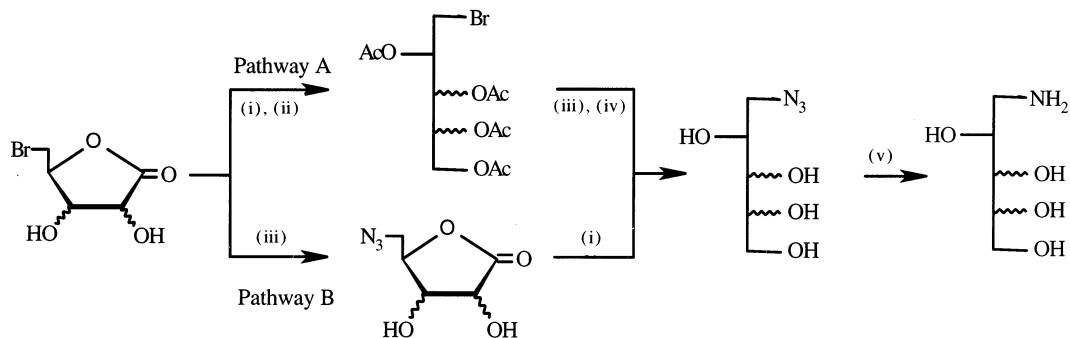
In the present work, we report two routes for the synthesis of novel 1-amino-1-deoxy-D- and L-pentitols, using 5-bromo-5-deoxy-D-pentono-1,4-lactones as starting material: either via 1-bromo-1-deoxy-D- and L-pentitols (Pathway A, Scheme 1), which are intermediates leading to potential glycosidase inhibitors and anti-HIV iminocyclitols [7–9], or via 5-azido-5-deoxy-D-pentono-1,4-lactones (Pathway B, Scheme 1).

We have previously described the preparation of 5-bromo-5-deoxy-D-pentono-1,4-lactones from unprotected D-pentono-1,4-lactones in good yields (75–95%) [10,11] by means of thionyl bromide in *N,N*-dimethylformamide.

Reduction of 5-bromo-5-deoxy-D-ribo-, 1,4-lactone (**1**) with sodium borohydride gave

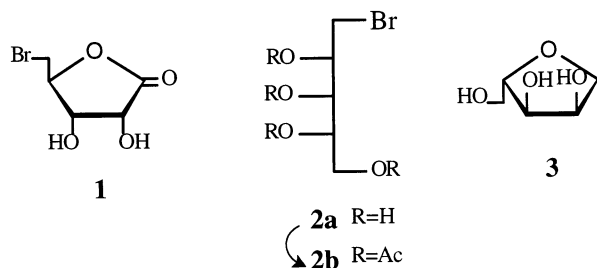
\* Corresponding author.

E-mail address: imane.stasik@sc.u-picardie.fr (I. Stasik)

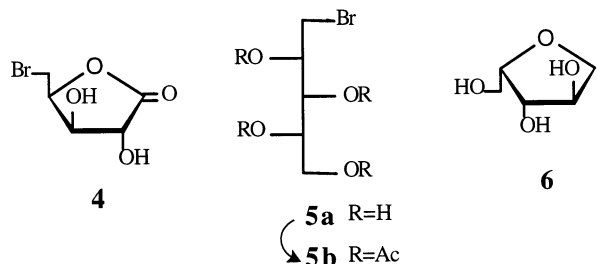


Scheme 1. (i)  $\text{NaBH}_4$ ,  $\text{H}_2\text{O}$ ,  $\text{EtOH}$ ; (ii)  $\text{Ac}_2\text{O}$ , pyr; (iii)  $\text{LiN}_3$ , DMF; (iv)  $\text{MeONa}$ ,  $\text{MeOH}$ ; (v)  $\text{HCOONH}_4$ ,  $\text{Pd-C}$ ,  $\text{EtOH}$ .

1-bromo-1-deoxy-L-ribitol (**2a**) (62%), with the anhydride **3** (38%) as by-product (Table 1). However, as compound **2a** is rapidly and quantitatively converted into anhydride **3**, we did not succeed in isolating it from the mixture of **2a** and **3**. To avoid any further intramolecular heterocyclisation, we treated the freshly prepared crude material with acetic anhydride in pyridine at room temperature and obtained, after purification, 2,3,4,5-tetra-*O*-acetyl-1-bromo-1-deoxy-L-ribitol (**2b**) in 60% yield.

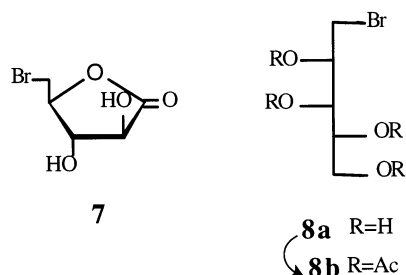


5-Bromo-5-deoxy-D-xylono-1,4-lactone (**4**) was treated under the same reduction conditions to afford 1-bromo-1-deoxy-L-xylitol (**5a**) (75%) and the anhydride **6** (25%). After acetylation of the crude material, **5b** was isolated in 70% yield.



Reduction of 5-bromo-5-deoxy-D-arabino-1,4-lactone (**7**) gave 1-bromo-1-deoxy-D-

lyxitol (**8a**) in quantitative yield. After acetylation, **8b** was isolated in 90% yield. In fact, it is well-known that halogenated D-arabinitol, in comparison with others halogenated pentitols, is less prone to cyclise to anhydro derivatives [12].



1-Azido-1-deoxypentitols were obtained by treatment of 1-bromo-1-deoxypentitols with lithium azide in *N,N*-dimethylformamide.

Thus, treatment of **2b** with lithium azide in *N,N*-dimethylformamide gave tetra-*O*-acetyl-1-azido-1-deoxy-L-ribitol (**9**) in quantitative

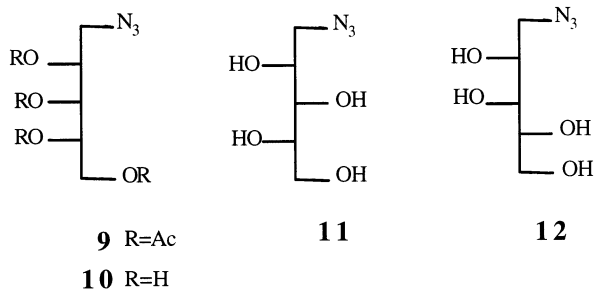
Table 1  
Reduction of 5-bromo-5-deoxy-D-pentono-1,4-lactone

Substrate	Product (% yield)
<b>1</b>	i) <b>2a</b> R=H (62) + <b>3</b> (38) ii) <b>2b</b> R=Ac (60)*
<b>4</b>	i) <b>5a</b> R=H (75) + <b>6</b> (25) ii) <b>5b</b> R=Ac (70)*
<b>7</b>	i) <b>8a</b> R=H (95) ii) <b>8b</b> R=Ac (90)*

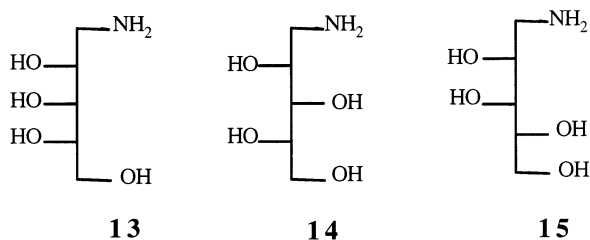
i)  $\text{NaBH}_4$ ,  $\text{H}_2\text{O-EtOH}$ ,  $0^\circ\text{C}$ . \* Isolated yield

yield. When reduction of **9** was performed, by catalytic hydrogen transfer (CHT) with ammonium formate as hydrogen donor and Pd–C as catalyst in ethanol, acetyl group migration occurred to give tri-*O*-acetyl-1-acetamido-1-deoxy-L-ribitol in 95% yield.

Deacetylation of **9** with sodium methoxide in methanol led to pure 1-azido-1-deoxy-L-ribitol (**10**), without further purification. In the same way, compounds **5b** and **8b** led quantitatively to 1-azido-1-deoxy-L-xylitol (**11**) and D-lyxitol (**12**), respectively.



Reduction of 1-azido-1-deoxy-L-ribitol was performed by CHT and afforded 1-amino-1-deoxy-L-ribitol (**13**) in quantitative yield. 1-Amino-1-deoxy-L-xylitol (**14**) and D-lyxitol (**15**) were also obtained quantitatively from the corresponding 1-azido-1-deoxypentitols.



To prevent anhydride formation, protection–deprotection steps and acetyl group migration, we have investigated a second route to aminopentitols via 5-azido-5-deoxy-D-pentono-1,4-lactones (Pathway B, Scheme 1).

5-Azido-5-deoxy-D-pentono-1,4-lactones were previously prepared in good yields by treatment of the corresponding 5-bromo-5-deoxy derivatives, with lithium azide in *N,N*-dimethylformamide [11].

5-Azido-5-deoxy-D-pentono-1,4-lactones were treated with sodium borohydride to give the corresponding 1-azido-1-deoxy-L or D-pentitols. When applied to 5-azido-5-deoxy-D-ribo-, D-xylono-, and D-arabinono-1,4-

Table 2

Product	<b>13</b> (%)	<b>14</b> (%)	<b>15</b> (%)
Pathway A	60	70	90
Pathway B	92	80	84

lactones this method yielded quantitatively 1-azido-1-deoxy-L-ribitol (**10**), L-xylitol (**11**) and D-lyxitol (**12**) respectively.

As previously reported, 1-amino-1-deoxy-L or D-pentitols **13**, **14** and **15** were obtained by CHT from the 1-azido-1-deoxy-L or D-pentitols **10**, **11** and **12** in quantitative yield.

The overall yields of both routes are reported in Table 2. Whilst Pathways A and B are both suitable for the synthesis of **14** and **15**, **13** is better prepared via Pathway B.

In summary, we have described two reliable routes for the synthesis of 1-amino-1-deoxy-L-ribitol (**13**), 1-amino-1-deoxy-L-xylitol (**14**) and 1-amino-1-deoxy-D-lyxitol (**15**). The major advantage of Pathway A is that it allows access to 1-bromo-1-deoxy-D- or L-pentitols, which are interesting intermediates.

### 3. Experimental

**General methods.**—<sup>1</sup>H and <sup>13</sup>C NMR spectra were recorded in CDCl<sub>3</sub>, D<sub>2</sub>O or C<sub>5</sub>D<sub>5</sub>N with Me<sub>4</sub>Si as internal standard on a Bruker 300 MHz spectrometer. Measurement of [α]<sub>D</sub> values was effected with a Jasco DIP-370 digital polarimeter, using a sodium lamp (λ = 589 nm), at 20 °C. Column chromatography was performed on Silica Gel (E. Merck 230–400 mesh). Analytical TLC was performed on E. Merck glass-backed Silica Gel sheets (Silica Gel F<sub>254</sub>).

**General procedure for reduction of 5-bromo- or 5-azido-5-deoxy-D-pentono-1,4-lactones.**—To a solution of 5-bromo- or 5-azido-5-deoxy-D-pentono-1,4-lactones (5.8 mM) in 2:1 water–EtOH (21 mL) was added ion-exchange resin (Amberlite IR-120H<sup>+</sup>, 3 mL). The mixture was cooled at 0 °C and stirred while NaBH<sub>4</sub> (1.6 equiv, 350 mg) was added at such a rate that the pH was maintained below 7. Then a further amount of NaBH<sub>4</sub> (1.9 equiv, 415 mg) was added to increase the

pH to 9. Stirring was continued at 0 °C for 1 h, before adding more ion-exchange resin (Dowex 50 × 8–100 ion) to decrease the pH to 3. The resin was then removed by filtration. The filtrate was concentrated and co-concentrated with MeOH (3 × 18 mL) to give the crude mixture, which was chromatographed on silica gel (4:1 EtOAc–MeOH). For the 1-bromo-1-deoxy-D- or L-pentitols, the crude material was treated with Ac<sub>2</sub>O (10 equiv) in pyridine (10 mL). Silica gel chromatography (9:1 and 7:3 petroleum ether–EtOAc) gave 2,3,4,5-tetra-O-acetyl-1-bromo-1-deoxy-D- or L-pentitols.

**2,3,4,5-Tetra-O-acetyl-1-bromo-1-deoxy-L-ribitol (2b).**—(1.33 g, 60%);  $[\alpha]_D - 3.1^\circ$  (*c* 1.27, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.02 (m, 1 H, H-1a), 4.22 (m, 1 H, H-1b), 5.11–5.17 (m, 2 H, H-2, H-4), 5.22 (m, 1 H, H-3), 3.33 (m, 1 H, H-5a), 3.52 (m, 1 H, H-5b), 1.93 (s, 3 H, CH<sub>3</sub>), 1.98 (s, 6 H, CH<sub>3</sub>), 2.01 (s, 3 H, CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  61.6 (C-1), 70.2, 69.4 (C-2; C-4), 70.5 (C-3), 29.8 (C-5), 20.5, 20.6 (CH<sub>3</sub>), 169.0, 169.5, 169.6, 170.3 (CO). Anal. Calcd for C<sub>13</sub>H<sub>19</sub>O<sub>8</sub>Br: C, 40.77%; H, 4.96%; Br, 20.86%. Found: C, 40.56; H, 5.01; Br, 20.81.

**2,3,4,5-Tetra-O-acetyl-1-bromo-1-deoxy-L-xylitol (5b).**—(1.55 g, 70%);  $[\alpha]_D - 8.7^\circ$  (*c* 1.26, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>):  $\delta$  3.96 (m, 1 H, H-1a), 4.16 (m, 1 H, H-1b), 5.05–5.15 (m, 2 H, H-2, H-4), 5.36 (m, 1 H, H-3); 3.27–3.34 (m, 2 H, H-5a, H-5b), 1.94, 1.95, 1.97, 2.0 (4s, 4 × 3 H, CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  61.6 (C-1), 69.0, 71.6 (C-2, C-4), 69.7 (C-3), 29.3 (C-5), 20.3, 20.4, 20.5, 20.7 (CH<sub>3</sub>), 169.4, 169.6, 169.7, 170.1 (CO). Anal. Calcd for C<sub>13</sub>H<sub>19</sub>BrO<sub>8</sub>: C, 40.77; H, 4.96; Br, 20.86. Found: C, 40.65; H, 4.90; Br, 20.84.

**2,3,4,5-Tetra-O-acetyl-1-bromo-1-deoxy-D-lyxitol (8b).**—(2.0 g, 90%);  $[\alpha]_D + 34.1^\circ$  (*c* 0.53, CHCl<sub>3</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  3.89 (dd, 1 H, *J*<sub>1a,2</sub> 6.9 Hz, H-1a), 4.21 (dd, 1 H, *J*<sub>1b,2</sub> 5.0 Hz, H-1b), 5.28–5.35 (m, 2 H, H-2, H-3), 5.12 (m, 1 H, *J*<sub>4,5a</sub> 5.7 Hz, H-4), 3.36 (dd, 1 H, H-5a), 3.50 (dd, 1 H, *J*<sub>4,5b</sub> 3.3 Hz, *J*<sub>5a,5b</sub> 11.6 Hz, H-5b), 1.98, 2.02, 2.04, 2.09 (4s, 4 × 3 H, CH<sub>3</sub>). <sup>13</sup>C NMR (75 MHz, CDCl<sub>3</sub>):  $\delta$  61.6 (C-1), 68.0; 70.0 (C-2; C-3), 68.6 (C-4), 30.8 (C-5), 20.5 (CH<sub>3</sub>), 169.4, 169.6, 169.9, 170.2 (CO). Anal. Calcd for

C<sub>13</sub>H<sub>19</sub>BrO<sub>8</sub>: C, 40.77; H, 4.96; Br, 20.86. Found: C, 40.92; H, 4.88; Br, 20.95.

**2,3,4,5-Tetra-O-acetyl-1-azido-1-deoxy-L-ribitol (9).**—(441.5 mg, 98%);  $[\alpha]_D - 4.6^\circ$  (*c* 0.88, CH<sub>2</sub>Cl<sub>2</sub>); <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub>):  $\delta$  4.24 (dd, 1 H, *J*<sub>1a,2</sub> 3.3 Hz, H-1a), 4.05 (dd, 1 H, *J*<sub>1a,1b</sub> 12.3 Hz, H-1b), 5.17 (m, 1 H, *J*<sub>1b,2</sub> 6.2 Hz, H-2), 5.24 (m, 1 H, *J*<sub>2,3</sub> 5.3 Hz, H-3), 5.10 (m, 1 H, *J*<sub>3,4</sub> 5.3 Hz, H-4), 3.44 (dd, 1 H, *J*<sub>4,5a</sub> 3.6 Hz, H-5a), 3.34 (dd, 1 H, *J*<sub>4,5b</sub> 6.8 Hz, *J*<sub>5a,5b</sub> 13.4 Hz, H-5b), 1.95 (s, 3 H, CH<sub>3</sub>), 2.01 (s, 6 H, CH<sub>3</sub>), 2.04 (s, 3 H, CH<sub>3</sub>); <sup>13</sup>C NMR (CDCl<sub>3</sub>):  $\delta$  61.6 (C-1), 69.4 (C-2), 69.6 (C-3), 70.3 (C-4), 50.1 (C-5), 20.5, 20.7 (CH<sub>3</sub>), 169.2, 169.7, 170.4 (CO). Anal. Calcd for C<sub>13</sub>H<sub>19</sub>N<sub>3</sub>O<sub>8</sub>: C, 45.25; H, 5.51; N, 12.17. Found: C, 45.10; H, 5.62; N, 12.40.

**1-Azido-1-deoxy-L-ribitol (10).**—(1.0 g, 98%); mp 59–61 °C;  $[\alpha]_D - 7.3^\circ$  (*c* 0.54, MeOH); <sup>1</sup>H NMR (300 MHz, pyr-*d*<sub>5</sub>):  $\delta$  4.30–4.44 (m, 3 H, H-1a, H-1b, H-2), 4.49 (m, 1 H, H-3), 4.66 (m, 1 H, H-4), 3.87–3.89 (m, 2 H, H-5a, H-5b). <sup>13</sup>C NMR (75 MHz, pyr-*d*<sub>5</sub>):  $\delta$  65.1 (C-1), 74.4 (C-2), 75.0 (C-3), 74.0 (C-4), 55.1 (C-5). Anal. Calcd for C<sub>5</sub>H<sub>11</sub>N<sub>3</sub>O<sub>4</sub>: C, 33.92; H, 6.21; N, 23.73. Found: C, 33.86; H, 6.35; N, 23.53.

**1-Azido-1-deoxy-L-xylitol (11).**—(985.6 mg, 96%);  $[\alpha]_D + 3.2^\circ$  (*c* 0.40, MeOH); <sup>1</sup>H NMR (300 MHz, pyr-*d*<sub>5</sub>):  $\delta$  3.98–4.03 (m, 3 H, H-1a, H-1b, H-3), 4.16 (m, 1 H, H-2), 4.22 (m, 1 H, *J*<sub>4,5a</sub> 4.2 Hz, *J*<sub>4,5b</sub> 7.5 Hz, H-4), 3.65 (dd, 1 H, H-5a), 3.56 (dd, 1 H, *J*<sub>5a,5b</sub> 12.8 Hz, H-5b). <sup>13</sup>C NMR (75 MHz, pyr-*d*<sub>5</sub>):  $\delta$  64.7 (C-1), 74.0 (C-2), 73.2 (C-3; C-4), 55.3 (C-5). Anal. Calcd for C<sub>5</sub>H<sub>11</sub>N<sub>3</sub>O<sub>4</sub>: C, 33.92; H, 6.21; N, 23.73. Found: C, 33.77; H, 6.11; N, 23.62.

**1-Azido-1-deoxy-D-lyxitol (12).**—(1.02 g, 99%); mp 73–75 °C;  $[\alpha]_D - 10.1^\circ$  (*c* 0.54, MeOH); <sup>1</sup>H NMR (300 MHz, pyr-*d*<sub>5</sub>):  $\delta$  4.29–4.33 (m, 3 H, H-1a, H-1b, H-3), 4.76 (m, 1 H, H-2), 4.62 (m, 1 H, H-4), 3.81 (dd, 1 H, *J*<sub>4,5a</sub> 6.8 Hz, H-5a), 3.94 (m, 1 H, *J*<sub>5a,5b</sub> 12.4 Hz, H-5b). <sup>13</sup>C NMR (75 MHz, pyr-*d*<sub>5</sub>):  $\delta$  65.2 (C-1), 71.9 (C-2), 73.2 (C-3), 72.3 (C-4), 56.1 (C-5). Anal. Calcd for C<sub>5</sub>H<sub>11</sub>N<sub>3</sub>O<sub>4</sub>: C, 33.92; H, 6.21; N, 23.73. Found: C, 33.91; H, 6.25; N, 23.80.

**General procedure for reduction of 1-azido-1-deoxy-D- or L-pentitols by catalytic hydrogen transfer.**—To a solution of 1-azido-1-deoxy-

D- or L-pentitols (200 mg) in EtOH (8 mL) was added, under an argon atmosphere, ammonium formate (5 equiv, 356 mg) and Pd–C (0.33 equiv). The reaction mixture was stirred at 70 °C for 1 h. After removal of the catalyst by filtration and evaporation of the solvent, the monoaminoalditol was obtained as the sole product.

*1-Amino-1-deoxy-L-ribitol* (**13**).—(167.2 mg, 98%);  $[\alpha]_D - 47.6^\circ$  (*c* 0.47, MeOH);  $^1\text{H}$  NMR (300 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  3.59–3.65 (m, 2 H, H-1a, H-3), 3.71–3.82 (m, 3 H,  $J_{4,5a}$  8.6 Hz, H-1b, H-2, H-4), 2.77 (dd, 1 H, H-5a), 2.96 (dd, 1 H,  $J_{4,5b}$  3.1 Hz,  $J_{5a,5b}$  13.4 Hz, H-5b).  $^{13}\text{C}$  NMR (75 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  64.8 (C-1), 74.3; 73.4 (C-2; C-4), 75.4 (C-3), 44.2 (C-5). Anal. Calcd for  $\text{C}_5\text{H}_{13}\text{NO}_4$ : C, 39.76; H, 8.61; N, 9.27. Found: C, 39.70; H, 8.70; N, 9.11.

*1-Amino-1-deoxy-L-xylitol* (**14**).—(167.2 mg, 98%);  $[\alpha]_D - 2.8^\circ$  (*c* 0.35, MeOH);  $^1\text{H}$  NMR (300 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  3.61–3.73 (m, 3 H, H-1a, H-1b, H-4), 3.79–3.89 (m, 2 H, H-2, H-3), 2.92–2.98 (m, 2 H, H-5a, H-5b).  $^{13}\text{C}$  NMR (75 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  64.7 (C-1), 74.2; 73.1 (C-2; C-3), 74.1 (C-4), 45.0 (C-5). Anal. Calcd for  $\text{C}_5\text{H}_{13}\text{NO}_4$ : C, 39.76; H, 8.61; N, 9.27. Found: C, 39.53; H, 8.33; N, 9.53.

*1-Amino-1-deoxy-D-lyxitol* (**15**).—(168.9 mg, 99%);  $[\alpha]_D - 8.2^\circ$  (*c* 1.79, MeOH);  $^1\text{H}$  NMR (300 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  3.60–3.63 (m, 2 H,

H-1a, H-1b), 3.87 (m, 1 H, H-2), 3.50 (q, 1 H,  $J_{2,3}$ ;  $J_{3,4}$  1.8; 8.1 Hz, H-3), 3.80 (m, 1 H,  $J_{4,5a}$  8.8 Hz, H-4), 2.89 (dd, 1 H, H-5a), 3.21 (dd,  $J_{4,5b}$  2.7 Hz,  $J_{5a,5b}$  13.1 Hz, H-5b).  $^{13}\text{C}$  NMR (75 MHz,  $\text{D}_2\text{O}$ ):  $\delta$  65.2 (C-1), 72.3 (C-2), 74.2 (C-3), 70.7 (C-4), 44.9 (C-5). Anal. Calcd for  $\text{C}_5\text{H}_{13}\text{NO}_4$ : C, 39.76; H, 8.61; N, 9.27. Found: C, 39.66; H, 8.17; N, 9.48.

## References

- [1] J.C. Norrild, C. Pedersen, J. Defaye, *Carbohydr. Res.*, 291 (1996) 85–98.
- [2] E.M. Acton, A.N. Fujiwawa, L. Goodman, D.W. Henry, *Carbohydr. Res.*, 33 (1974) 135–151.
- [3] J. Cleophax, J. Defaye, S.D. Gero, *Bull. Soc. Chim. Fr.*, (1967) 104–107.
- [4] W. Lai, O.R. Martin, *Carbohydr. Res.*, 250 (1993) 185–193.
- [5] H. Dietrich, R.R. Schmidt, *Carbohydr. Res.*, 250 (1993) 161–176.
- [6] P.M. Myerscough, A.J. Fairbanks, A.H. Jones, I. Bruce, S.S. Choi, G.W.J. Fleet, S.S. Al-Daher, I.C. di Bello, B. Winchester, *Tetrahedron*, 48 (1992) 10177–10190.
- [7] B. Winchester, G.W.J. Fleet, *Glycobiology*, 2 (1992) 199–210.
- [8] I. Lundt, R. Madsen, S. Al-Daher, B. Winchester, *Tetrahedron*, 50 (1994) 7513–7520.
- [9] X. Qian, F.M. Varas, C.H. Wong, *Bioorg. Med. Chem. Lett.*, 6 (1996) 1117–1122.
- [10] V. Bouchez, I. Stasik, D. Beaupère, R. Uzan, *Carbohydr. Res.*, 300 (1997) 139–142.
- [11] V. Bouchez, I. Stasik, D. Beaupère, R. Uzan, *Tetrahedron Lett.*, 38 (1997) 7733–7736.
- [12] M. Benazza, M. Massoui, R. Uzan, G. Demailly, *Carbohydr. Res.*, 275 (1995) 421–431.